# Principles of purification of macromolecules

## Genetics 222 Method and Logic in Experimental Genetics

Reference

Protein Purification: Principles and Practice Robert K. Scopes Third Edition Springer-Verlag 1994

# **Problem**

- Often need one component in a cell in purified form
  - why purified?
  - what does "pure" mean?
  - sometimes, partial enrichment is enough
- Task is to separate desired component from a complex mixture
- It's usually important to maintain the activity of the component throughout the process
  - yield is important
  - but so is intactness, activity
  - achieving one is often at the expense of the other

#### • Components can be:

- nucleic acids (DNA, RNA)
- proteins
- protein/nucleic acid complexes (snp's, transcription complexes)
- large cellular complexes (ribosomes, spliceosomes)

# **Protein purification**

#### Proteins

- huge number of proteins (> than the # of genes encoded in a genome)
- most cell types in multicellular organisms express tens of thousands of different proteins
- relative abundances of various proteins vary widely

#### Goals of protein purification

- obtain a particular protein free of others and other cell components
- obtain a good yield (absolute amount and proportion of starting amount)
- maintain the activity of the protein
  - Problems:
    - denaturation
    - proteolysis
    - in vitro mixing
- a measurement of how well the process worked
- some characterization of the purified protein

## 1. Develop a quantitative assay

- quantitative measurement of activity
- quantitative immunoassay
- gel electrophoresis assay (immunoblot or gel activity assay)
- generally want to increase <u>Specific Activity</u> at each step

#### **Definition of Specific Activity**

- for proteins: units per milligram of total protein
- "unit" is defined by the researcher, and can be different when described by different people or different venders
- a unit is a quantitative measure of activity, usually associated with a turnover rate (for enzymes) or amount needed for stoichiometric binding (for receptors, ligands, DNA binding proteins)
- theoretical maximum for any given protein

## 2. Obtain source of material

- whole organisms
- organs or tissues
- embryos
- tissue culture cells
- microorganisms
- need to understand and then weigh +'s and -'s of any particular source, e.g.:
  - how hard is it to obtain, grow, handle
  - amount of proteolytic activity
  - may sometimes be better to use a lower producing source that is cleaner
  - is the protein active in a particular source?
  - are inhibitors present in a particular source?
  - some organs and tissues have connective tissues that are hard to remove
- overproduction in a heterologous system (basis for much of biotechnology industry)

# **Protein purification**

### Low abundance





# **Protein purification**

## **High abundance**





## 3. Make an extract from source

- almost always want to keep it very cold (just above freezing)
- •gentle breaking
- in some cases can obtain big purification in one step by separating cellular compartments (e.g., purify nuclei from cytoplasm before extracting nuclear proteins)

## 4. Begin separating components

- remove nucleic acids, polysaccharides, cell membrane debris
- ammonium sulfate precipitations, other crude fractionations (pH or other salt precipitations, antibody clearing, "autolysis")
- these crude steps are often needed to avoid ruining or overloading chromatography agents

## 5. Fractionate by chromatography

- several steps are almost always needed
- need to assay for amount and purity at each step
- need a way to decide when you're finished

# **Separation principles**

## Sizing

- principle is based on exclusion of larger molecules from pores in resin; smaller molecules require longer times of transit
- sizing resins = "gel filtration"
- different matrices have different size ranges
- examples: Sephadex, Sephacryl, Ultrogel, some HPLC resins

## Ion exchange

- separate on the basis of net charge
- wash and elute with higher salt concentrations
- determine highest [salt] that your protein binds to the resin, and use this to load or to pre-wash before eluting your protein
- examples: DEAE cellulose, phosphocellulose, some HPLC resins

## **Affinity chromatography**

- separate based on binding to residues specific or semi-specific to the protein you are purifying
- easy to overload with non-specific proteins
- examples: sequence-specific DNA sites, antibody resin

## **Purification table**

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1		1
1		1
		1
1		1
		-

# **Purification of DNA**

#### • DNA

- many types you might want to purify (genomic, plasmid, viral, particular segment of genome, some linear, some circular)
- pretty hardy, so fairly harsh methods can be used
- specificity of desired segment is relatively easy to assess

#### • Assays for purification of DNA molecules

- functional assays
  - complementation in a microorganism or tissue culture cell (usually must be mutant, or missing)
  - Inhibition of function in a microorganism or tissue culture cell
- hybridization
  - must already know something about the DNA sequence
  - blots, in situ hybridization, microarray hybridization
- PCR
  - especially helpful for gene families, related sequences
- DNA sequencing
  - particularly possible now because of huge databases from genome sequencing projects

# **Purification of RNA**

#### Assays

- why? mRNA purification or enrichment, snp's, enzymatic RNAs, structural RNAs
- not as hardy as DNA, although easier to work with than proteins if special care is taken to avoid RNases
- slight chemical difference between RNA and DNA makes biochemical properties quite different (don't heat with Mg+2)
- most RNAs are single stranded, but with much secondary (double stranded) character

#### Assays for purification of RNA molecules

- cDNA and EST libraries
  - screen by PCR, hybridization, direct sequencing
- gel assays (Northern blotting)
- in situ hybridization
- functional assays
  - microinjection
  - **inhibition**

## **Food for Thought**

# What are the pitfalls/caveats of purification approaches?